

Guanidine/Guanidinium Nitrate; a Mild and Selective *O*-Deacetylation Reagent that Leaves the *N*-Troc Group Intact.

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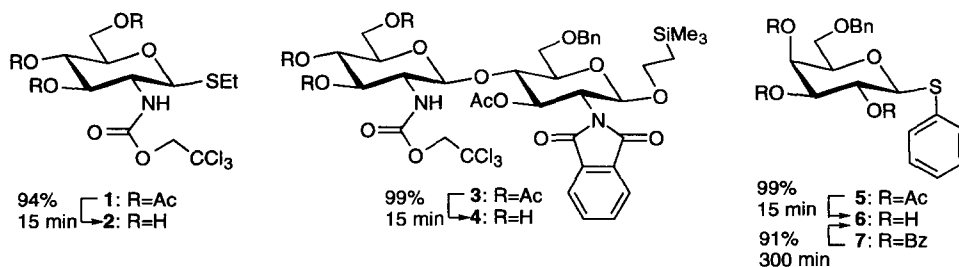
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Abstract. Treatment of *O*-acetyl-protected sugars with a methanolic solution of guanidine/guanidinium nitrate caused the removal of the acetyl groups (91-99% isolated yield), without affecting other protecting groups. Removal of *O*-benzoyl groups required a longer reaction time. Of special merit is the stability of the 2,2,2-trichloroethoxycarbonylamino (*N*-Troc) group under these weakly basic reaction conditions.
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Protection of the amino group of 2-deoxy-2-aminosugars by the 2,2,2-trichloroethoxycarbonyl (Troc) group to give an *N*-Troc carbamate, is a good alternative to e.g. phthalimides, both during glycoside synthesis and subsequent deprotection.¹⁻⁵ The *N*-Troc group is relatively stable under acidic conditions, which permits removal of *O*-acetyl groups by methanolic HCl (e.g. **1**→**2**).⁶ However, attempted selective *O*-deacetylation of **1** under basic conditions (MeONa/MeOH, EtONa/EtOH, *t*BuOK/*t*BuOH, MeONa/MeOH/H₂O, NH₃/MeOH, KCN/EtOH, K₂CO₃/MeOH/H₂O) was unsuccessful; the *N*-Troc group was transformed into the corresponding carbamate.

Kunesch introduced guanidine (liberated from guanidinium chloride by the addition of one equivalent of sodium ethoxide) as an efficient *O*-deacetylation reagent.⁷ Later, Wong used guanidinium carbonate for the same purpose.⁸ Attempted *O*-deacetylation of **1** by the Kunesch procedure failed to spare the *N*-Troc group and the corresponding carbamate was formed. However, when the amount of MeONa was lowered to 0.2 equivalents, clean *O*-deacetylation of **1** gave **2** in 94 % yield, without any cleavage of the *N*-Troc group (Chart 1). Similarly, the disaccharide **3** was *O*-deacetylated to give **4** in 99% yield; the acetyl group in the 3-position was unaffected, probably due to steric hindrance by the *N*-Phth group. The guanidine/guanidinium

Chart 1.



nitrate reagent (G/GHNO₃) was investigated further with a number of *O*-acylated sugar derivatives, carrying various additional protecting groups. Both *S*- and *O*-glycosides were unaffected by the G/GHNO₃ reagent. In addition, a number of standard protecting groups (NPhth, PhCHO₂, BnO, Me₂CO₂, Ph₂^tBuSiO) were equally stable. The differences in steric hindrance by these groups was demonstrated by the varying reaction times (15-120 min) needed for full *O*-deacetylation. In contrast to the rapid (15 min) *O*-deacetylation of **5** (→**6**, 99%), *O*-debenzoylation of **7** (→**6**, 91%) required a reaction time of 300 min. The average isolated yield of *O*-deacylated products was 97%, using ten differently protected sugars. The tetrachlorophthalimido, *N*-Fmoc, and *O*-Troc protecting groups were unstable against the G/GHNO₃ reagent.

General O-deacylation procedure. A clear stock solution of the G/GHNO₃ reagent was prepared by dissolving guanidinium nitrate (622 mg, 5 mmol) in MeOH/CH₂Cl₂ (50 mL, 9:1) and adding methanolic MeONa (1 mL, 1 M). The stock solution was kept at room temperature for several weeks without any observed decrease in activity.

The *O*-acylated sugar derivative (0.1 mmol) was dissolved in an aliquot of the stock solution (6 mL). The reaction was monitored by TLC (SiO₂, heptane/EtOAc 1:1 and CH₂Cl₂/EtOH 10:1). When the *O*-deacylation was complete, the mixture was neutralized by addition of Amberlite IR 120 H⁺ and the solvent was removed. The residue was chromatographed (SiO₂, CH₂Cl₂/EtOH 10:1) to give the *O*-deacylated saccharide (e.g. Chart 1). The *O*-deacylated compounds were *O*-acetylated to recreate the starting materials **1**, **3**, and **5**. The identity of all new compounds was shown by NMR and HRMS.

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